

# 3M™ Sponge Stick and 3M™ Hydrated Sponge

How 3M meets your organization's sample collection needs.



## Quality

- Lot-to-lot quality release testing
- Certificate of Analysis available for every lot
- Continuous improvement to exceed 3M technical standards

## Consistency

- Process controlled manufacturing
- Robust supplier standards and controls

## Global coverage and support

- Supply chain—climate controlled products
- Technical support
- Global coverage with local support

## USDA-FSIS

- Validated 3M Sponges for food contact and environmental swab samples (FSIS Directive 3/28/13)

### Key Considerations

### Facts

Removal of bacterial contaminants from the surface

The release of these bacteria from the swab/sponge for quantitative measure  
Subsequent cultivation

Cellulose and Polyurethane sponges are proven to be equivalent for sampling environmental surfaces\*+.

Free of biocides

Biocide-free cellulose sponge maintains viability for wide range of organisms. *Listeria* can be maintained for 72 hours of refrigeration\*.

Toxicity and environmental friendliness

Cellulose sponges are made from renewable biomass Polyurethane sponges are made from reaction of polyols, diisocyanates, catalysts, and additives.

Strength and durability

3M sponges made with cellulose are tested for withstanding scrubbing on multiple surfaces\*.

Batch to batch consistency

3M sponges made with cellulose raw materials are sorted during inspection process so that the chemical and mechanical properties of the cellulose sponges are consistent from batch to batch\*.

Ambient storage temperature

- Lethen is stable at ambient storage temperature for up to 2 months\*.
- NB is stable at ambient storage temperature\*.

# Cellulose and Polyurethane sponges are proven to be equivalent for sampling environmental surfaces.

## Key Considerations

- The effectiveness of the swabbing technique depends on the efficacy of these three individual components:
  - The removal of bacterial contaminants from the surface
  - The release of these bacteria from the swab/sponge for quantitative measurements
  - Their subsequent cultivation
- To optimize the potential for consistent, accurate laboratory results all batches of sponges should be tested for sterility and efficacy to ensure every product is of consistent quality.
- The guidelines in the Microbiology Laboratory Guidebook (MLG) of USDA and Bacteriological Analytical Manual (BAM) of FDA specifies sponge composition to be non-bactericidal, cellulose or polyurethane as necessary for environmental sampling <sup>(1,2)</sup>.
  - Cellulose sponges are manufactured using natural ingredients, cellulose derived from wood pulp, sodium sulphate and hemp fibers.
  - Polyurethane sponges are made by reacting a polyol, a type of complex alcohol, with diisocyanate in the presence of suitable catalysts and additives.

## Publications

Recent scientific publications by FDA and academia evaluate sponge performances and demonstrate outcomes consistent with 3M internal studies:

1. Sheth, I., *et.al.* (2018) Comparison of three enrichment schemes for the detection of low levels of desiccation-stressed *Listeria* spp. from select environmental surfaces. *Food Control*, 84; 493-498
  - FDA results showed no statistically significant difference on swabbing *Listeria* spp. from stainless steel surface between sponges made from cellulose (SSL100, 3M) and polyurethane (EZ-10DE-PUR, World Bioproducts).
2. Keeratipibul, S., *et.al.* (2017) Effect of swabbing techniques on the efficiency of bacterial recovery from food contact surfaces. *Food Control*, 77; 139-144
  - Cellulose sponge and polyurethane (PU) foam swabs provided a greater swab efficiency on biofilm recovery among different surface types and microorganisms.
  - Statistically significant high values of biofilm swabbing efficiency are in bold.
3. Internal 3M studies demonstrate that cellulose and polyurethane sponges do not show statistically significant differences in their pick up and release efficiencies from stainless steel surface.

**Table 3: Number of positive samples by each sponge sampler material (Manufacturer) after sampling**

| Sponges material     | Cellulose (3M) | Cellulose (Nasco) | Polyurethane (Worldbioproduct) |
|----------------------|----------------|-------------------|--------------------------------|
| Positive control (5) | 5              | 5                 | 5                              |
| Negative control (5) | 0              | 0                 | 0                              |
| Samples (20)         | 10             | 8                 | 13                             |

**Table 2: Biofilm swab efficiency of each swab type.**

| Bacteria                | Surface Type | Biofilm Swab Efficiency* of Each Swab Type |                     |                    |                     |
|-------------------------|--------------|--|---------------------|--------------------|---------------------|
|                         |              | Cotton                                     | Gauze               | PU Foam            | Cellulose Sponge    |
| <i>E. coli</i>          | Stainless    | 47.8 ± 0.8c                                | <b>51.4 ± 0.7a</b>  | 48.3 ± 0.4b,c      | <b>51.3 ± 0.1a</b>  |
|                         | New PSU      | 50.1 ± 0.7c                                | <b>52.0 ± 0.6ab</b> | <b>52.6 ± 0.9a</b> | 51.0 ± 1.1bc        |
|                         | Old PSU      | 49.7 ± 0.5b                                | 49.6 ± 1.0ab        | 49.7 ± 1.0ab       | <b>50.0 ± 0.8a</b>  |
| <i>S. aureus</i>        | Stainless    | 49.4 ± 0.2d                                | 54.2 ± 0.7b         | 53.4 ± 0.1c        | <b>55.0 ± 0.6a</b>  |
|                         | New PSU      | 48.9 ± 0.2d                                | <b>52.6 ± 0.5ab</b> | 51.3 ± 0.2c        | <b>53.6 ± 0.1a</b>  |
|                         | Old PSU      | 47.5 ± 0.1d                                | 52.0 ± 0.1d         | 50.5 ± 0.2c        | <b>52.8 ± 0.3a</b>  |
| <i>S. Typhimurium</i>   | Stainless    | 46.7 ± 0.7c                                | 47.0 ± 0.7bc        | <b>50.0 ± 0.4a</b> | <b>48.5 ± 0.3ab</b> |
|                         | New PSU      | 46.2 ± 0.7d                                | 51.9 ± 1.7b         | <b>55.2 ± 0.1a</b> | 51.6 ± 1.7bc        |
|                         | Old PSU      | 45.1 ± 0.4c                                | 44.9 ± 1.0c         | <b>49.3 ± 0.7a</b> | 47.7 ± 2.0b         |
| <i>L. monocytogenes</i> | Stainless    | 48.2 ± 0.1c                                | 50.0 ± 0.2b         | 50.2 ± 0.0b        | <b>51.0 ± 0.1a</b>  |
|                         | New PSU      | 47.8 ± 0.1c                                | <b>52.5 ± 0.3a</b>  | 50.8 ± 0.1b        | <b>52.9 ± 0.1a</b>  |
|                         | Old PSU      | 48.2 ± 0.1c                                | 49.8 ± 0.2b         | 50.4 ± 0.1b        | <b>51.7 ± 0.1a</b>  |
| <b>Total Average</b>    |              | 48.0                                       | 50.7                | 51.0               | 51.4                |

\*Data are means ± SD for three determinations. Means in the same row with no letters in common are significantly different (P<0.05).

1. Microbiology Laboratory Guidebook, 8.10. Isolation and Identification of *Listeria monocytogenes* from Red Meat, Poultry and Egg Products, Ready-To-Eat Siluriformes (Fish) and Environmental Samples. Revision 10. (2017).

2. Bacteriological Analytical Manual, Chapter 10. Detection and Enumeration of *Listeria monocytogenes* in Foods. (2015). U.S Food and Drug Administration.



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